

0040-4039(94)02076-0

Synthesis and Antiretroviral Evaluation of 3-Alkyl 2-Piperazinone Nucleoside Analogs

Abdellah Benjahad^(a), Rachida Benhaddou^(a), Robert Granet^(a), Mourad Kaouadji^(a), Pierre Krausz^{(a)*}, Salomon Piekarski^(a), François Thomasson^(b), Claudine Bosgiraud^(c) and Sylvie Delebassée^(c)

^aLaboratoire de Chimie des Substances Naturelles, Faculté des Sciences, 123, Avenue Albert Thomas
87060 Limoges Cedex, France

^bService commun de RMN, UFR Pharmacie, Domaine de La Merci 38706 La Tronche Cedex, France

^cUFR de Pharmacie, 2 rue du Docteur Marcland, 87025 Limoges Cedex, France

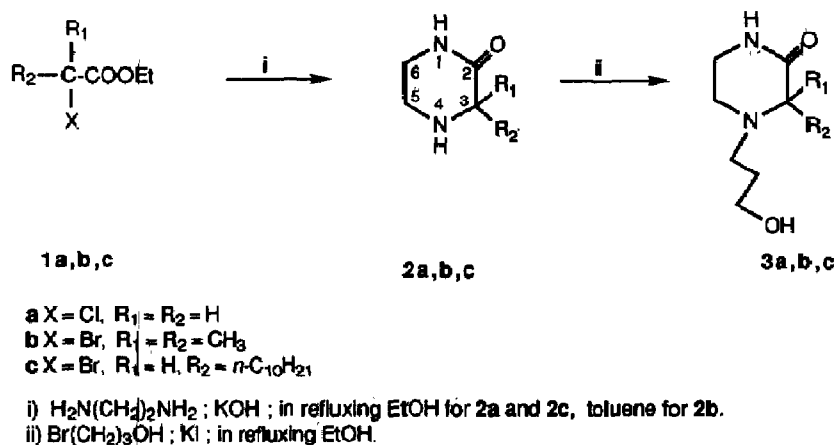
Abstract: Glycosylation of 3-alkyl *N*⁴-(3-hydroxypropyl) 2-piperazinones by protected 1-*O*-acetyl ribofuranoses produces nucleoside analogs in which the base is separated from the sugar by a hydrocarbon spacer arm. The preliminary *in vitro* test results against retroviruses seem promising for compounds bearing a long alkyl chain.

As part of our research program on the synthesis of nucleoside analogs¹⁻³, we became interested in the preparation of such analogs derived from 2-piperazinone in which the "base" is linked to the ribose by an aliphatic chain. It is now known that an aliphatic chain not only serves as a neutral linkage but also affects the coiling of DNA. Increasing the length of the spacer arm linkage would thus enhance bending of DNA, leading to base pair opening where the bases then become susceptible to attack by reactive groups.⁴ Spacer arms have also been used in oligomer nucleoside antisense synthesis for preventing the enzymatic degradation by nucleases.⁵ Also, a hydrophobic tail was introduced on the base to improve cellular uptake.⁶ We wish to report herein the synthesis and characterisation of a new class of nucleoside analogs, and preliminary results of their *in vitro* activity against sheep retroviruses. One of the representative compounds of this new series shows moderate activity. The synthetic route used in this work is illustrated in Schemes I and II.

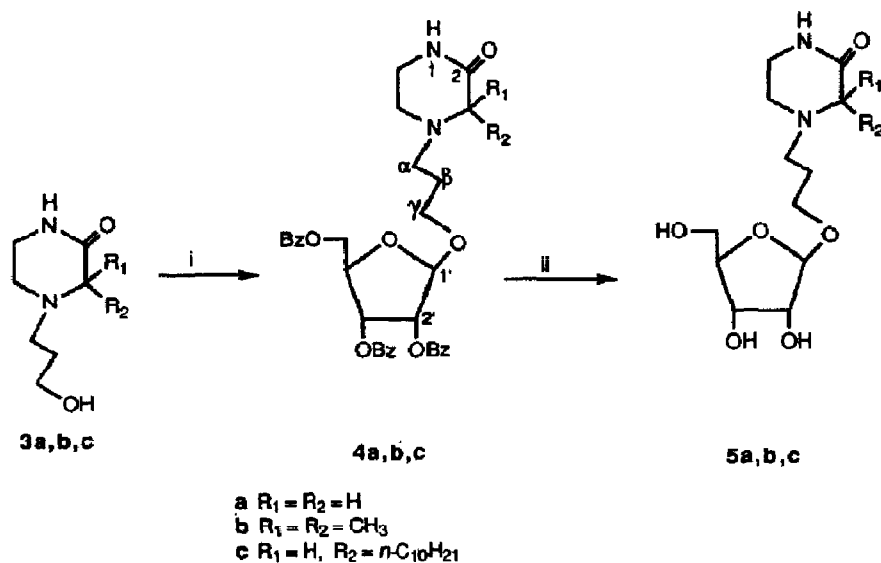
Starting piperazinones **2a,b,c** were prepared by a modification of known methods.⁷⁻⁹ Syntheses were carried out by the action of ethylenediamine (6 eq) on the corresponding ethyl α -halogenoalkanoate **1a,b,c** followed by neutralization and then cyclisation. After the usual work-up, the expected compounds were obtained with 50, 62 and 80% yields, respectively. By refluxing **2a,b,c** in dry ethanol with 3-bromopropanol (1 eq) and a catalytic amount of KI for 24 h, **3a,b,c** were obtained after recrystallization in EtOH in acceptable yields.¹⁰ The *N*-4 linkage between *n*-propanol and piperazinone in compounds **3a,b,c** was shown in the corresponding ¹H NMR spectra by both the disappearance of the amine proton previously recorded at δ 1.82 for **2a**, δ 1.68 for **2b** and δ 1.61 for **2c** as well as the continuous presence of the amide proton at δ 6.10 for **3a**, δ 6.18 for **3b** and δ 6.21 for **3c**.

Glycosylation of **3a,b,c** was performed according to the method of Hannessian.¹¹ Use of dichloroethane as solvent even in the presence of a large excess of SnCl₄ resulted in poor yields (15%) probably due to the precipitation of a complex formed from the base and SnCl₄. Inversely, in acetonitrile, higher yields were obtained, as no precipitation occurred in the presence of excess SnCl₄. In a typical experiment, under argon, 2 mmol of SnCl₄ were added to a solution of 1-*O*-acetyl 2,3,5-tri-*O*-benzoyl- β -D ribofuranose (1 mmol) in dry acetonitrile. After 15 min, 1 eq of each aglycon **3a,b,c** was introduced in the medium and the solution was stirred for 3h at room temperature. The resulting solution was then neutralized by NaHCO₃ and extracted with

chloroform. After separation by column chromatography on silica gel (eluent $\text{CHCl}_3\text{-EtOH} = 99:1$) compounds **4a,b,c** were obtained with 50, 45 and 41% overall yields respectively.¹² Compound **4c** gave a chromatographically unseparable mixture of diastereoisomers due to the asymmetrical C-3 atom. Subsequent attempts to separate the diastereoisomers by various chromatographic means remain presently unsuccessful.



Scheme I



Scheme II

For all new compounds **4a,b,c** satisfactory ^1H , ^{13}C NMR and mass spectra data were obtained on chromatographically homogeneous samples. As an example we have included the ^1H and ^{13}C NMR data for compound **4a**.¹³ In all glycosides, the small coupling constant value ($J_{1',2'} < 1$ Hz) strongly indicates a β configuration of the ribofuranose moiety; this is also confirmed by the ^{13}C NMR data relative to the anomeric carbon (δ 105.5-105.7).

Treatment of **4a,b,c** with ammonia in methanol at room temperature for one week gave deprotected products **5a,b,c** in good yields.^{14,15}

Compounds **2b**, **3b**, **5b** and **5c** were evaluated for their antiviral activities against a mammalian retrovirus, the Visna virus, strain K796, in sheep choroid plexus cells (SCP cells). AZT, the most potent inhibitor of this virus was also tested in order to compare the effectiveness of the newly synthesized compounds. Antiviral activities were measured by a modified MTT¹⁶ method applied to adherent cells.¹⁷ SCP cells were infected with Visna virus at a MOI of 0.5 and incubated in the presence of the tested compound. After 5 days incubation, the number of viable cells was determined. The results are shown in Table 1.

Compounds	EC50 (μM) ^a	CC50 (μM) ^b
2b	-	>700
3b	-	>400
5a	-	>500
5b	-	>800
5c	80	170
AZT	5	>500

^aEC50: 50% antiviral effective concentration.

^bCC50: 50% cytotoxic concentration.

TABLE 1

As shown in Table 1, among the compounds tested, piperazinone **2b** and the intermediate **3b** showed no antiviral activities. Final piperazinone nucleosides modified with a spacer arm exhibited different results according to R_1 and R_2 . Only compound **5c** with an aliphatic long chain had moderate antiviral activity; however, this molecule was cytotoxic at a concentration greater than 170 μM . No comparable inhibition such as that observed with AZT was obtained. An extensive evaluation is in progress and complete results will be published in due course.

In summary, we have successfully synthesized and tested new nucleosides with a spacer arm linking the sugar and the base. New compounds in this series are currently under investigation in our laboratory.

Acknowledgments.

We are pleased to acknowledge J.M. Davis for her help in the preparation of the English version of the manuscript, Mrs. Meraville and Dr. Couquet (Laboratoire Départemental d'Analyse de la Haute Vienne) for recording the mass spectra. We also thank ANVAR (Association Nationale de Valorisation et d'Aide à la Recherche) for their financial support.

REFERENCES:

1. Depelley, J.; Granet, R.; Krausz, P.; Piekarski, S.; Bosgiraud, C.; Beaussoleil, S. *Nucleosides and Nucleotides*, **1994**, *13* (4), 1007-1010.
2. Beaussoleil, S.; Bosgiraud, C.; Depelley, J.; Granet, R.; Krausz, P.; Nicolas, J.A.; Piekarski, S. French patent N° 9202401 (28/02/92).
3. Czernecki, S.; Ezzitouni, A.; Krausz, P. *Synthesis*, **1990**, *8*, 651-653.
4. Dodin, G.; Kühnel, S. M.; Demersman, P.; Kotzyba, J. *Anti Cancer Drug Design*, **1993**, *8*, 361-368.
5. Caufield, T. J.; Prasad, C. V. C.; Deleki, D. J.; Pronty, C. P.; Saha, A. K.; Upson, D. A.; Kruse, L. I. *Bioorg. & Medicinal Chem. Lett.*, **1994**, *4*, 1497-1500.
6. Sagi, J.; Szemzo, A.; Ebinger, K.; Szabolcs, A.; Sagi, G.; Ruff, E.; Otvos, L. *Tetrahedron Lett.*, **1993**, *34*, 2191-2194.
7. Aspinall, S. R. *J. Am. Chem. Soc.*, **1940**, *62*, 1202-1204.
8. Shigemi, K.; Hideyo, K. *Yakugaku Zasshi*, **1962**, *82*, 909-912.
9. Piekarski, S. *Oléagineux*, **1962**, *10*, 785-789.
10. **3a** : yield 81%; syrup; IEMS *m/z* : 159 [M+H]⁺, 113 [M-(CH₂)₂OH]⁺; **3b**: yield 60%; mp 96-98°C; IEMS *m/z* : 186 [M]⁺, 171 [M-CH₃]⁺; **3c**: yield 52%; syrup; IEMS *m/z* : 298 [M]⁺, 157 [M-(C₁₀H₂₁)]⁺.
11. Hanessian, S.; Banoub, J. *Methods in Carbohydrate Chemistry*, Academic Press: New York, 1980; Vol III, pp 243-245.
12. **4a** : syrup; *R_f* = 0.32 (CHCl₃-EtOH = 95:5); [α]²⁰_D = +23.9° (c 1.96, CHCl₃); **4b** : syrup; *R_f* = 0.41 (CHCl₃-EtOH = 95:5); [α]²⁰_D = +18.2° (c 1.24, CHCl₃); **4c** : syrup; *R_f* = 0.48 (CHCl₃-EtOH = 95:5).
13. **4a** : ¹H NMR (200 MHz, CDCl₃, δ ppm) *base*: 3.09 (2H, s, H-3), 2.61 (2H, t, *J* = 6.1 Hz, H-5), 3.33 (2H, dt, *J* = 6.2, 2.2 Hz, H-6), 5.68 (1H, br s, NH); *arm*: 2.47 (2H, t, *J* = 6.9 Hz, α-H), 1.73 (2H, quintuplet, *J* = 6.6 Hz, β-H), 3.55 (1H, dt, *J* = 9.4, 6.2 Hz, γ_a-H), 3.84 (1H, dt, *J* = 9.6, 6.1 Hz, γ_b-H); *ose*: 5.25 (1H, br s, H-1'), 5.66 (1H, dd, *J* = 4.9, 0.6 Hz, H-2'), 5.85 (1H, dd, *J* = 6.5, 4.9 Hz, H-3'), 4.71 (2H, m, H-4', H-5'_a), 4.54 (1H, dd, *J* = 12.7, 6.7 Hz, H-5'_b); *benzoyl groups*: 7.90, 8.04, 8.08 (2H each, dd, *J* = 8.5, 1.4 Hz, H-2,6), 7.35 (6H, m, H-3,5), 7.53 (6H, m, H-4). ¹³C NMR (50 MHz, CDCl₃, δ ppm) *base*: 169.4 (C-2), 49.1 (C-3), 54.0 (C-5), 56.7 (C-6); *arm*: 41.4 (C-α), 26.7 (C-β), 66.2 (C-γ); *ose*: 105.6 (C-1'), 75.5 (C-2'), 72.6 (C-3'), 78.9 (C-4'), 65.0 (C-5'); *benzoyl groups* : 129.0 (3C, C-1), 129.7 (6C, C-2,6), 128.4 (4C, C-3,5), 128.5 (2C, C-3,5), 133.2, 133.4 and 133.5 (1C each, C-4), 165.3, 165.4 and 166.1 (1C each, C-7).
14. **5a** : yield 85%; syrup; *R_f* = 0.52 (MeOH - 30% NH₄OH = 99 : 1); [α]²⁰_D = -30.2° (c 0.55, MeOH); **5b** : yield 75%; syrup; *R_f* = 0.62 (MeOH - 30% NH₄OH = 99 : 1); [α]²⁰_D = -21.1° (c 0.40, MeOH); **5c** : yield 72%; syrup; *R_f* = 0.68 (MeOH - 30% NH₄OH = 99 : 1).
15. Each reaction was followed by TLC on silica gel.
16. Pauwels, R.; Balzarini, J.; Baba, M.; Snoeck, R.; Schols, D.; Herdewijn, P.; Desmyter, J.; De Clercq, E. *J. Virol. Meth.*, **1988**, *20*, 309-321.
17. Bosgiraud, C.; Delebassée, S. to be published.

(Received in France 27 September 1994; accepted 17 October 1994)